# Clinical utility of detecting HPV in urine samples as primary cervical cancer screening including HPV 16/18 genotyping

Radha Rani Padhy, M.D.

PGY – 4, OBGYN Staten Island University Hospital Staten Island, NY, USA



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases



#### No financial relationships or conflict of interest to disclose



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases



# **Cervical Cancer Screening – Statistics**

#### Table 71 (page 1 of 5). Use of Pap smears among women aged 18 and over, by selected characteristics: United States, selected years 1987–2015

Excel and PDF versions (with more data years and standard errors when available): http://www.cdc.gov/nchs/hus/contents2016.htm#071.

[Data are based on household interviews of a sample of the civilian noninstitutionalized population]

Characteristic	1987	1993	1999	2000	2005	2008	2010	2013	2015
		Per	cent of wor	nen having	a Pap sme	ar within th	e past 3 yea	ars <sup>1</sup>	
18 years and over, age-adjusted <sup>2,3</sup> 18 years and over, crude <sup>2</sup>	74.1 74.4	77.7 77.7	80.8 80.8	81.3 81.2	77.9 77.7	75.6 75.1	73.7 73.2	70.4 69.4	70.2 69.0
Age								<b>-</b>	
18-44 years   18-20 years     18-20 years   21-44 years     21-24 years   25-44 years     25-44 years   45-64 years     45-54 years   55-64 years     65 years and over   65-74 years     75 years and over   75 years and over	83.3 59.4 86.1 85.3 86.3 70.5 75.7 65.2 50.8 57.9 40.4	84.6 66.8 86.2 86.1 86.3 77.2 82.1 70.6 57.6 64.7 48.0	86.8 65.3 89.2 85.3 89.9 81.7 83.8 78.4 61.0 70.0 50.8	84.9 59.8 87.8 84.1 88.5 84.6 86.3 82.0 64.5 71.6 56.7	83.6 61.1 86.3 84.0 86.8 80.6 83.4 76.8 54.9 66.3 42.7	81.8 57.5 84.8 80.2 85.7 78.8 81.0 76.0 50.0 61.6 37.5	80.4 52.0 84.0 81.1 84.6 76.9 79.9 73.2 47.1 58.0 34.6	77.2 38.6 81.6 74.6 83.2 73.9 78.6 68.6 42.7 54.5 27.9	76.1 34.0 81.1 69.7 83.5 75.5 79.7 71.1 42.3 52.9 28.1

• https://www.cdc.gov/nchs/data/hus/2016/071.pdf: Accessed Jan 15, 2018 (1)

• A large proportion of the population is **LOST TO SCREENING PROGRAMS**, either due to poor resources, cultural barriers or avoidance of a pelvic exam.



## Current screening guidelines:

Population	Recommended Screening Method	Comment
Women younger than 21 years	No screening	
Women aged 21–29 years	Cytology alone every 3 years	
Women aged 30–65 years	Human papillomavirus and cytology cotesting (preferred) every 5 years Cytology alone (acceptable) every 3 years	Screening by HPV testing alone is not recommended*
Women older than 65 years	No screening is necessary after adequate negative prior screening results	Women with a history of CIN 2, CIN 3, or adenocarcinoma in situ should continue routine age-based screening for a total of 20 years after spontaneous regression or appropriate management of CIN 2, CIN 3, or adenocarcinoma in situ
Women who underwent total hysterectomy	No screening is necessary	Applies to women without a cervix and without a history of CIN 2, CIN 3, adenocarcinoma in situ, or cancer in the past 20 years
Women vaccinated against HPV	Follow age-specific recommendations (same as unvaccinated women)	

 Pap smears every 3 years starting at age 21 or co-testing with pap smears and HPV every 5 years starting at age 30 <sup>(2).</sup>



# Primary hrHPV screening

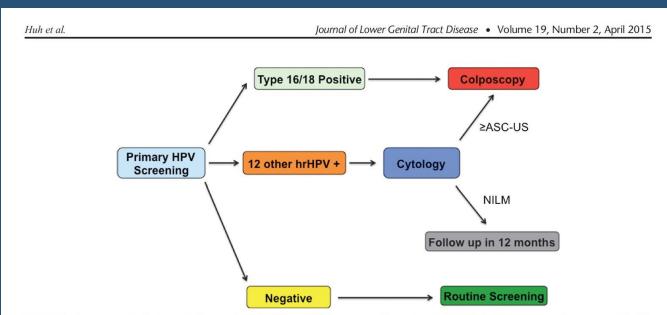


FIGURE 1. Recommended primary HPV screening algorithm. HPV, human papillomavirus; hrHPV, high-risk human papillomavirus; ASC-US, atypical squamous cells of undetermined significance; NILM, negative for intraepithelial lesion or malignancy.

 An interim clinical guideline was published in 2015 on primary HPV screening, which can be considered as an alternative to cytology alone, co-testing, and other current US cytology-based cervical cancer screening approaches <sup>(3).</sup>

• For women > 25 years: Routine screening: every 3 years <sup>(3).</sup>



# Objective

 To detect high-risk human papillomavirus (hrHPV) mRNA in urine samples and compare their concordance with hrHPV mRNA in cervical samples including HPV 16 and 18/45 genotyping

#### Primary endpoint:

 Comparing hrHPV detection in cervical samples to urine samples including HPV 16 and HPV 18/45 genotyping

#### <u>Secondary endpoint</u>

 Determining the positive predictive value (PPV) for urine hrHPV detection for high grade histologic lesions (≥ CIN 2)

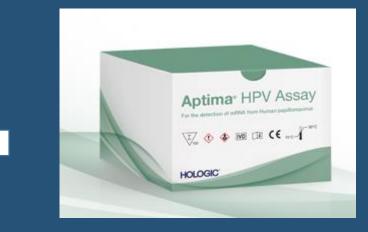


# Methods

#### Panther Hologic system



• <u>Aptima HPV assay</u>



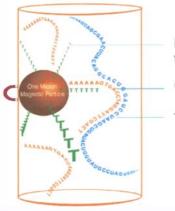
• Transcription-mediated amplification (TMA) system using reverse transcriptase  Detects (qualitative E6/E7 mRNA) hrHPV types: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68.
(Does not discriminate between the 14 high-risk types)



# **Testing Methodology**

#### **<u>1. Target capture</u>**

 Isolates and purifies the target HPV mRNA by use of capture oligomers that are linked to magnetic micro-particles <sup>(4).</sup>



Poly-T oligomer bound to magnetic particle Capture oligonucleotide

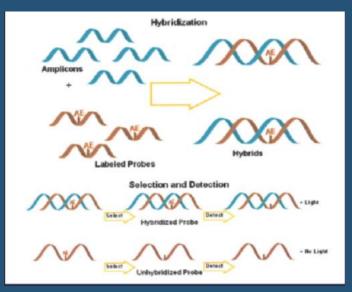
Target sequence

#### 2. Transcription-<u>Mediated</u> <u>Amplification:</u>

 Uses MMLV reverse transcriptase and T7 RNA polymerase to generate multiple copies of RNA amplicon from the DNA copy template <sup>(4).</sup>

#### **<u>3. Hybrid Protection Assay:</u>**

 Hybridizes the amplicon to single stranded nucleic acids with chemiluminescent labels that are complimentary to the amplicon <sup>(4).</sup>.





Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

# hrHPV Genotyping

Urine samples which were positive for hrHPV further underwent hrHPV genotyping using the Aptima HPV 16, 18/45 Genotype Assay (Detects E6/E7 HPV mRNA)



• Can differentiate HPV 16 from HPV 18 and/or HPV 45, but **does not** differentiate between HPV 18 and HPV 45.



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

#### Power

- A sample size of 186 patients was calculated to estimate a 95% confidence interval (CI) for the sensitivity of the urinary HPV test for detecting HPV
  - (assuming  $80\% \pm 8\%$  of HPV positive patients will have positive test and the prevalence of HPV in the study population is 50%.)
- This equals to 62 patients per group.
  - With this sample size, the **precision for specificity will be 90%±6%.**
- Results for **189 patients** are presented so far.



## Recruitment

#### • Location:

 Centre for Women's Health at Staten Island University Hospital, NY.

#### Urine collection:

• During clinic visits

#### • Inclusion criteria:

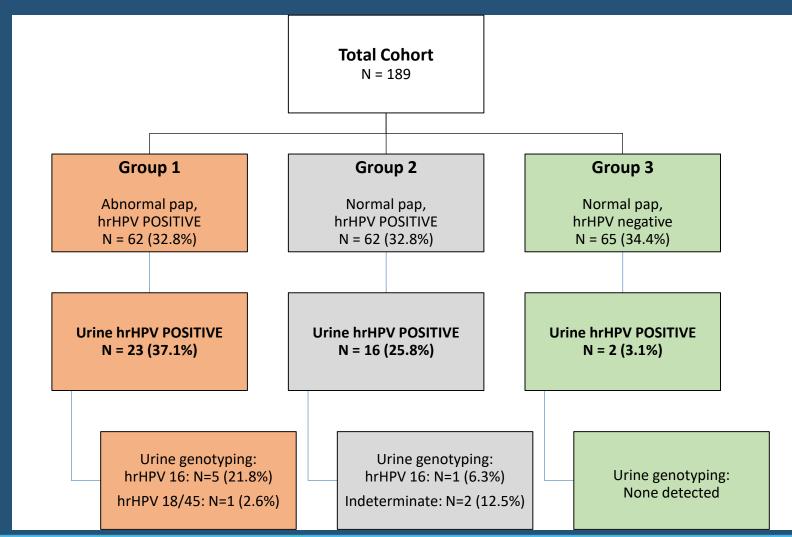
- Age 25 or above
- Pap smear performed within past 360 days

#### Exclusion criteria

- Pregnant at time of urine collection
- History of HPV vaccine administered

#### Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

## Study Flowsheet





Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

# Total Cohort Characteristics

					oup 1 = rmal pap,	Group 2 = Normal pap,		Group 3 = Normal pap,		
		٦	otal	hrHPV	hrHPV POSITIVE		hrHPV POSITIVE		hrHPV negative	
		Ν	%	N	%	N	%	N	%	
Ν		189		62	32.80%	62	32.80%	65	34.39%	
Age (years)										
	Median	41		36		42		45		
Age Groups (years)										
	<30	24	12.70%	15	24.19%	7	12.28%	2	3.08%	
	≥30-49	108	57.14%	34	62.96%	35	61.40%	39	60.00%	
	≥50	57	30.16%	13	24.07%	20	35.09%	24	36.92%	0.006
Race										
	Hispanic	92	48.68%	29	53.70%	40	70.18%	23	35.38%	
	Non-hispanic	97	51.32%	33	61.11%	22	38.60%	42	64.62%	0.004
Parity										
	0-1	47	24.87%	22	40.74%	8	14.04%	17	26.15%	
	2-4	117	61.90%	34	62.96%	42	73.68%	41	63.08%	
	≥5	15	7.94%	3	5.56%	6	10.53%	6	9.23%	0.080
Smoking										
	Yes	30	15.87%	12	22.22%	10	17.54%	8	12.31%	
	No	159	84.13%	50	92.59%	52	91.23%	57	87.69%	0.553
Hx of past or present sexually transmitted disease (STD)										
	Yes	28	14.81%	14	25.93%	9	15.79%	5	7.69%	
	No	161	85.19%	48	88.89%	53	92.98%	60	92.31%	0.061

**ASCCP**2018 Annual Meeting



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

# Urine hrHPV detection (compared to cervical hrHPV detection)

	<u>Cervical</u>		
	hrHPV	hrHPV	
	negative	positive	Total
<u>Urine samples</u>			
hrHPV positive	2	39	41
hrHPV negative	63	85	148
Total	65	124	189

				Overall
	Cohen's			percent
	kappa	95% CI	p value	agraamant
1	карра	95% CI	p value	agreement

Prevalence	(95% CI)	Sensitivity	(95% CI)	Specificity	(95% CI)	PPV	(95% CI)	NPV (95% CI)
65.6%	(58.3-72.3%)	31.5%	(23.6-40.5)	96.9%	(88.4 – 99.5%)	95.1%	(82.2-99.1%)	42.6% (34.6-51.0%)



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

## Urine hrHPV detection (normal versus abnormal pap smear with cervical hrHPV positivity)

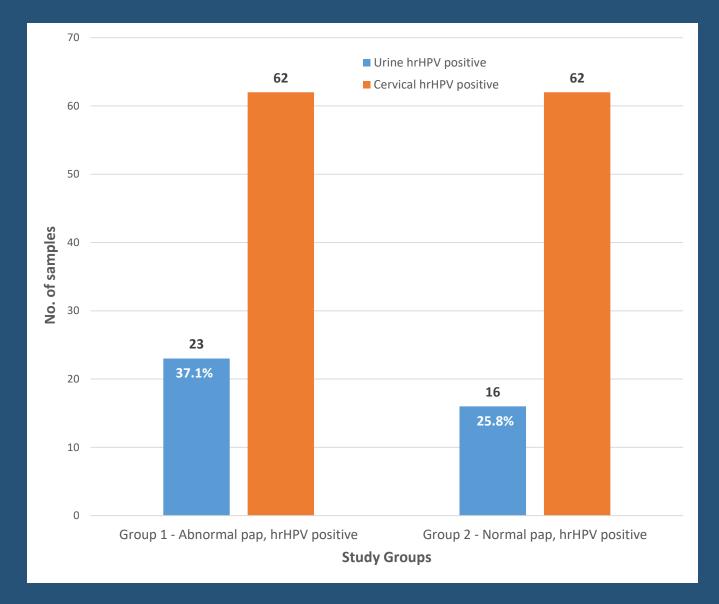
	<u>Group 1 –</u> Abnormal pap, hrHPV positive	<u>Group 2 –</u> Normal pap, hrHPV positive	Total
Urine hrHPV positive	23	16	49
Urine hrHPV negative	39	46	85
Total	62	62	124

Prevalence	e (95% CI)	Sensitivity	(95% CI)	Specificity	(95% CI)	PPV	(95% CI)	NPV (95% CI)
50.0%	(38.7-57.8%)	37.1%	(25.4-50.3%)	74.2%	(61.3 – 84.1%)	60.0%	(42.2-74.0%)	54.1% (43.0-64.9%)



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

 A higher proportion of patients with abnormal cytology and hrHPV positivity had positive urine samples (37.1%; 23/62) versus patients with normal cytology and hrHPV positivity (25.8%; 16/62).





## Urine hrHPV prevalence in Group 1 according to cervical cytology

	Urine hrHPV positive	Cervical hrHPV positive
ASCUS	8	25
ASC-H	4	7
LGSIL	10	23
HGSIL	1	7
Total	23	62



p = 0.64



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

# Urine hrHPV detection for high grade cervical histology (≥ CIN2)

	<u>Cervical</u>	Cervical colposcopy/LEEP biopsies			of sam.	20				18
	≤ CIN 1	≥ CIN 2	Total		<b>2</b> 0	36.4%			14	18
<u>Urine samples</u>				_	10	-	-		77.8%	
hrHPV positive	20	14	34		0					
hrHPV negative	55	18	73			≤ CII		ervical Hi		N 2
Total	75	32	107	p = 0.64		Cervical Histology				
$Drevelence (05% Cl) \qquad Sensitivity (05% Cl)$				עמט /		<b>CI</b> )				

60

50

40

ples

55

Prevalence (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)	PPV (95% CI)	NPV (95% CI)
29.9% (21.6-39.6%)	43.8% (26.8-62.1%)	73.3% (61.7 – 82.6%)	41.2% (25.1-59.2%)	<mark>75.3%</mark> (63.7-84.4%)



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

#### ASCCP2018 Annual Meeting

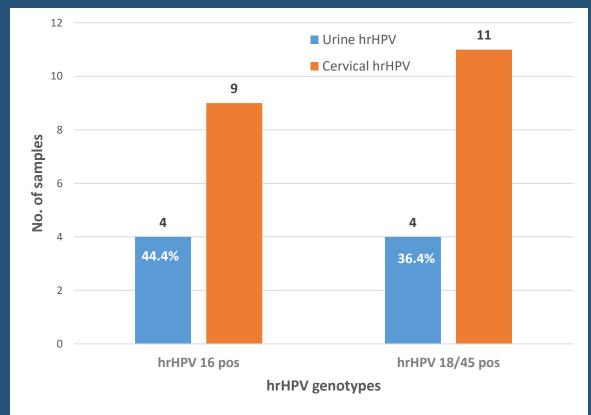
Urine hrHPV positive

Cervical hrHPV positive

# hrHPV genotyping in urine and cervical samples

Genotyping was performed on 57 cervical samples, of which 20 (35.1%) were either hrHPV 16 positive or hrHPV 18/45 positive

	Pap smear hrHPV		Urine hrHPV positive	
hrHPV 16 pos	9	15.79%	4	44.44%
hrHPV 18/45 pos	11	19.30%	4	36.36%





Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

## Urine hrHPV Genotype results

Cervical samples				Urine samp	oles	Cytology	Colposcopy / LEEP	Median time (days) from pap collection to urine collection
hrHPV	HPV 16	HPV 18/45	hrHPV	HPV 16	HPV 18/45			
pos	pos	neg	pos	Detected	Not detected	ASC-H	CIN 3	16
pos	pos	neg	pos	Detected	Not detected	HGSIL	CIN 3	48
pos	pos	neg	pos	Detected	Not detected	NILM	CIN 3	146
pos	neg	pos	pos	Not detected	Detected	LGSIL	CIN 2	0
pos	Not done	Not done	pos	Detected	Not detected	LGSIL	CIN 3	354
pos	Not done	Not done	pos	Detected	Not detected	ASCUS	CIN 2	187
pos	Not done	Not done	pos	Detected	Not detected	LGSIL	CIN 2	116

• The **genotype-specific concordance** for HPV 16 and 18/45 was **100%** for all the urine samples when compared to the cervical samples.



# Percent agreement of hrHPV genotyping between urine and cervical samples

	<u>Cervical</u>		
	HPV 16 or 18/45	HPV 16 or 18/45	
	positive	negative	Total
Urine samples			
hrHPV 16 or 18/45			
positive	8	0	8
hrHPV 16 or 18/45			
negative	12	37	49
Total	20	37	57

Overall agreement	<mark>78.9%</mark>
Positive percent agreement	<mark>40.0%</mark>





## Paired cervical and urine samples

		iroup 1 = ap, hrHPV POSITIVE	Group 2 = Normal pap, hrHPV POSITIVE		
Ν	10		6		
Urine hrHPV positive	4	40.0%	2	33.3%	
Genotyping – URINE samples	2	20.0%	1	16.7%	
		hrHPV 18/45 pos x 1, hrHPV 16 pos x 1		hrHPV 16 pos	
	1= ASCUS, hrHPV				
Cytology and HPV status of cervical	2= LGSIL, hrHPV p	os (16 neg, 18 pos)			
samples corresponding to urine	3= LGSIL, hrHPV p	os (16 neg, 18 pos)	1 = NILM, HPV pos (16 pos, 18 neg)		
hrHPV positive samples	4= ASC-H, hrHPV <sub>F</sub>	pos (16 pos, 18 neg)	2 = NILM, HPV pos (16 pos, 18 neg)		

**ASCCP**2018 Annual Meeting

• The positive percent agreement for both groups is **37.5%** 



## Discussion

- In this study, the overall sensitivity of urine hrHPV detection is low at 31.5%, however the specificity and PPV are above 90% (96.9% and 95.1%) respectively.
- There is a **FAIR agreement** between cervical and urine samples (k=0.22, Cl 0.14-0.30, p=0.04) with an overall percent agreement of 54%.
- Median time of pap collection to urine collection was similar in both Groups 1 and 2 who had hrHPV positivity in cervical samples (122 and 124 days respectively).



### Discussion

- A higher percentage of patients with ≥ CIN 2 histology had positive urine HPV results (77.8%; 14/18) versus patients with < CIN 2 histology (36.4%; 20/55).
- The specificity was **73.3%** with a sensitivity of 43.8%
- The PPV was **41.2%**





## Discussion

- For HPV 16 or HPV 18/45 genotyping, the overall agreement was 78.9% with a positive percent agreement of 40.0%.
  - However, of all the cervical hrHPV positive samples, **less than half** were HPV 16 or HPV 18/45 positive.
  - The genotype-specific concordance was **100%**
- In the group of paired cervical and urine samples, the **positive percent agreement was 37.5%.** 
  - This is slightly higher than the positive percent agreement for Groups 1 and 2 (positive cervical hrHPV) at **31.5%**



# Current Challenges with urine HPV detection

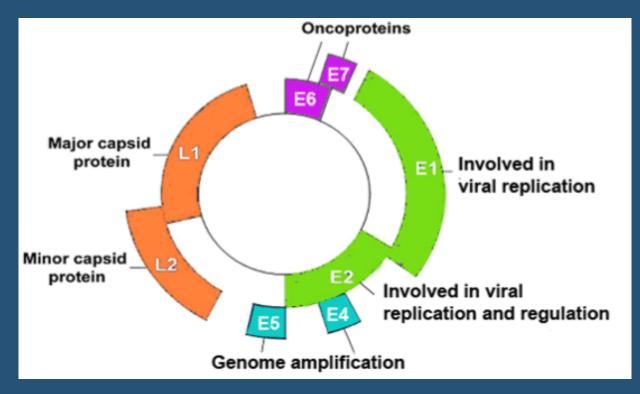
- There is <u>no consensus</u> of mode of urine collection (i.e. storage, amount, type of void) and HPV DNA extraction from these samples.
- Some studies have found higher sensitivity and specificity for DNA hrHPV detection in first void samples <sup>(5).</sup>
- Senkomago et al (2016):
  - "... no difference on HR-HPV detection for first void, initial stream and midstream urine for unfractionated and pellet fractions"<sup>(6)</sup>



## **HPV DNA Detection in Urine Samples**

• Most HPV DNA tests are based on the PCR detection of the L1 gene (7-8).

High sensitivity, BUT low specificity <sup>(7)</sup>





Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

#### ASCCP2018 Annual Meeting

# Summary of Studies (Worldwide)

Year	Author	Country	Sample size	HPV Detection Test	Sample collection	Urine HPV positive specimens	k agreement (Overall agreement %)	For	Sensitivity (%)	Specificity (%)
2014	Ducancelle et al (9)	France		INNO-LiPA HPV genotyping assay	paired urine / cervical samples	98	0.80 (90%)	n/a	n/a	n/a
2014	Combita et al (10)	Columbia		E7-MPG, IARC multiplex genotyping assay	paired urine / cervical samples	343	0.81 (79%)	19 HPV genotypes	91	74
2014	Bernal et al (11)	Seville	125		paired urine / cervical samples	66	0.76 (88%)	14 HPV genotypes	91	90
2015	Stanczuk et al (12)	Scotland	5318	Cobas 4800 test	paired urine / cervical samples	578	0.9 (n/a)	CIN 2+	63	90
2015	Stanczuk et al (13)	UK	100	Cobas 4800 test	paired urine / cervical samples	78	, , , , , , , , , , , , , , , , , , ,	CIN 2+ CIN 3+	80 91	23 27
2015	Lim et al (5)	Korea	100	Roche HPV assay, Abbott HPV assay	paired urine / cervical samples	Roche - 56 Abbot - 69	n/a	HPV 16/18	Roche - 79 Abbott - 82	Roche - 100 Abbott - 100
2015	Burroni et al (14)	Italy		INNO-LiPA HPV genotyping Extra	paired urine / cervical samples	51	n/a ( <sup>80%</sup> )	n/a	n/a	n/a
2015	Hagihara et al (15)	Japan		Anyplex TM II HPV 28 detection kit	paired urine / cervical samples	98	0.79 (98%)	19 HPV genotypes	68	100
2016	Khunamornpong et al (16)	Thailand	123	Cobas 4800 test	paired urine / cervical samples	30	· · · ·	14 hrHPV genotypes	69	93



# Summary of studies (Worldwide)

Year	Author	Country	Sample size	HPV Detection Test	Sample collection	Urine HPV positive specimens	k agreement (Overall agreement %)	For	Sensitivity (%)	Specificity (%)
2017	Cuzick et al (17)	UK	501	Trovagene HPV test	paired urine / cervical samples	396		CIN 3+ CIN 2+	91 81	(For <cin -="" 2)="" 25<="" td=""></cin>
	Nilyanimit et al (18)	Thailand	164	HPV GenoArray Assay	paired urine / cervical samples	53	,	21 HPV genotypes	57	71
2017	Lee et al (8)	Korea	-	Direct PCR-free colorimetric detection	paired urine / cervical samples	12	n/a	n/a	n/a	n/a
2017	Tshomo et al (19)	Bhutan	89	PCR E7-MPG, G5+/6+	paired urine / cervical samples	PCR E7-MPG - 56 G5+/6+ - 27	,	21 HPV genotypes	PCR E7-MPG - 80 G5+/6+ - 58	PCR E7-MPG - 61 G5+/6+ - 89
2017	Vergara et al (20)	Chile	543	PCR-RLB	paired urine / cervical samples	301	, ,	30 HPV genotypes	82	94
2017	Leeman et al (21)	Netherlands		SPF10-DEIA-LIPA25 assay, GP5+/6+ EIA- LMNX	paired urine / cervical samples	SPF10 - 67,70 GP5+/6+ - 60, 60	SPF10 - 0.83,0.86 (n/a) GP5+/6+ - 0.75,0.78 (n/a)	CIN2+	SPF10 - 95-100 GP5+/6+ - 95	SPF10 - 29-32 GP5+/6+ - 42
2018	Lorenz et al (22)	Brazil		HPV-HR (PCR based assay)	paired urine / cervical samples	259	. ,	CIN 2+ CIN 3+	83 86	51 36



# Summary of Studies: USA

Year	Author	Country	Sample Size	HPV detection Kit	Sample Collection	Urine HPV positive specimens (N)	k agreement (Overall agreement %)	For	Sensitivity (%)	Specificity (%)
	Sahasrabuddhe et al (23)	USA		genotyping test assay	paired urine/ cervical samples	54	0.55 (79%)	CIN 2/3	81	53
	Sahasrabuddhe et al (24)	USA		linear array HPV	paired urine/ cervical samples	Trovagene - 56 LA-HPV - 47	0.65 (92%)	CIN 2/3 CIN 3 CIN 2/3 CIN 3	Trovagene - 92 Trovagene -100 LA-HPV - 81 LA-HPV - 90	Trovagene - 29 Trovagene - 25 LA-HPV - 42 LA-HPV - 38
2014	Mendez et al (25)	USA		genotyping assay	paired urine/ cervical samples	22	n/a (76%)	n/a	n/a	n/a
2015	Senkomago et al (6)	USA	37	Ŭ	paired urine/ cervical samples	21-27	n/a	CIN 2	90	35
2016	Piyathilake et al (26)	USA		method	paired urine/ cervical samples	269		37 HPV genotypes	n/a	n/a



# HPV mRNA Detection in Cervical Samples

- E6/E7 mRNA has been studied in cervical samples with overall conclusions of **increased specificity** compared to HPV DNA assays <sup>(7, 27,29, 30).</sup>
- Theory:
  - Increased expression of HPV E6/E7 oncoproteins supports cervical tumorigenesis <sup>(7).</sup>

- In active infections, the hallmark is production of mRNA and proteins (7).
- mRNA can be detected by **reverse transcriptase** <sup>(7).</sup>



## **APTIMA HPV mRNA detection**

- Asciutto et al (Sweden, 2018): APTIMA mRNA based hrHPV testing on self collected urine and vaginal samples compared to clinician-taken cervical samples <sup>(28)</sup>
- For Urine Samples:
  - Overall:
  - Compared to this study:
  - For detecting HSIL/AIS/cancer:
  - For detecting ≥ CIN2 pathology:

Sensitivity: 48.1%, Specificity: 82.8% <sup>(28)</sup> Sensitivity: 31.5%, Specificity: 96.9%

Sensitivity: 44.8%, Specificity: 61.9% <sup>(28)</sup> Sensitivity: 43.8%, Specificity: 73.3%



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

## Conclusion

- The utility of mRNA reverse-transcriptase mediated amplification methodology using the APTIMA HPV assay is <u>suboptima</u>l in detection of hrHPV mRNA in urine samples and therefore cannot be considered for primary cervical cancer screening.
- However, due to its <u>high specificity</u>, mRNA urine HPV detection can be utilized in disease surveillance for patients with ≥ CIN 2 histology.







• Power of study

- Small sample size of paired cervical and urine samples
- Comparison of urine HPV detection in distinct groups with and without cytologic abnormality
- All hrHPV positive genotypes were not tested for on cervical and urine samples





## **Future Direction**

• Standardizing mode of urine collection and storage

• Obtaining larger sample size of paired urine and cervical samples

• HPV mRNA detection in patients with ≥ CIN 2 histology

• Using broad HPV genotyping assays with current methodology.



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

# Acknowledgements

#### Department of OBGYN

• Adi Davidov, MD (Director of Gynecology)

#### Department of Research

- Louise Madrigal, RN
- Gina Alcide, RN
- Meagan Sills, Administrative Director
- Department of Medicine
  - Almir Spahiu









Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases

#### References

- 1. Centers for Disease Control and Prevention. Use of Pap smears among women aged 18 and over, by selected characteristics: United States, selected years 1987-2015. https://www.cdc.gov/nchs/data/hus/2016/071.pdf. Accessed January 15, 2018.
- 2. Massad LS, Einstein MH, Huh WK, Katki HA, Kinney WK, Schiffman M, Solomon D, Wentzensen N, Lawson HW. 2012 ASCCP Consensus Guidelines Conference. 2012 updated consensus guidelines for the management of abnormal cervical cancer screening tests and cancer precursors. J Low Genit Tract Dis. 2013 Apr;17(5 Suppl 1):S1-S27.
- 3. Huh WK, Ault KA, Chelmow D, Davey DD, Goulart RA, Garcia FA, Kinney WK, Massad LS, Mayeaux EJ, Saslow D, Schiffman M, Wentzensen N, Lawson HW, Einstein MH. Use of primary high-risk human papillomavirus testing for cervical cancer screening: interim clinical guidance. Gynecol Oncol. 2015 Feb;136(2):178-82.
- 4. Aptima HPV 16 18/45 Genotype Assay Package Insert. Hologic 2007-2015
- 5. Lim, M. C., Lee, D., Hwang, S., Hwang, N. R., Lee, B., Shin, H. Y., . . . Joo, J. (2017). Comparison of the Abbott RealTime High Risk HPV test and the Roche cobas 4800 HPV test using urine samples. *Journal of Virological Methods, 243*, 74-79. doi:10.1016/j.jviromet.2017.01.020
- 6. Senkomago, V., Marais, A. D., Rahangdale, L., Vibat, C., Erlander, M., & Smith, J. (2016). Comparison of urine specimen collection times and testing fractions for the detection of high-risk human papillomavirus and high-grade cervical precancer. *Journal of Clinical Virology*, 74, 26-31. doi:10.1016/j.jcv.2015.11.005
- Chambers, G., Millan, D., Cuschieri, K., Cubie, H. A., & Graham, S. V. (2013). Assessing the detection of human papillomavirus late mRNA in liquid base cytology samples for risk stratification of cervical disease. *Journal of Medical Virology, 86*(4), 627-633. doi:10.1002/jmv.23793
- 8. Khan, F. H., N. H., & Malik, H. I. Proteins and Peptides Re-emergence in Prophylactics and Therapeutics. In *Advances in protein chemistry*. Romania: OMICS International. doi:https://www.esciencecentral.org/ebooks/ebookchapter/proteins-and-peptides-re-emergence-in-prophylactics-and-therapeutics" title="Proteins and Peptides Re-emergence in Prophylactics and Therapeutics" Proteins and Peptides Re-emergence in Prophylactics and Therapeutics" and Therapeutics" Proteins and Peptides Re-emergence in Prophylactics and Therapeutics" Proteins and Peptides Re-emergence in Prophylactics and Therapeutics
- 9. Ducancelle, A., Legrand, M. C., Pivert, A., Veillon, P., Guillou-Guillemette, H. L., Brux, M. A., ... Payan, C. (2014). Interest of Human Papillomavirus DNA quantification and genotyping in paired cervical and urine samples to detect cervical lesions. Archives of Gynecology and Obstetrics. doi:10.1007/s00404-014-3191-y
- 10. Combita, A. L., Gheit, T., Gonzalez, P., Puerto, D., Murillo, R. H., Montoya, L., . . . Wiesner, C. (2016). Comparison between Urine and Cervical Samples for HPV DNA Detection and Typing in Young Women in Colombia. Cancer Prevention Research, 9(9), 766-771. doi:10.1158/1940-6207.capr-16-003
- 11. Bernal, S., Palomares, J. C., Artura, A., Parra, M., Cabezas, J. L., Robles, A., & Mazuelos, E. M. (2014). Comparison of urine and cervical samples for detecting human papillomavirus (HPV) with the Cobas 4800 HPV test. Journal of Clinical Virology, 61(4), 548-552. doi:10.1016/j.jcv.2014.10.001
- 12. Stanczuk, G., Baxter, G., Currie, H., Lawrence, J., Cuschieri, K., Wilson, A., & Arbyn, M. (2016). Clinical validation of hrHPV testing on vaginal and urine self-samples in primary cervical screening (cross-sectional results from the Papillomavirus Dumfries and Galloway—PaVDaG study). *BMJ Open, 6*(4). doi:10.1136/bmjopen-2015-010660
- 13. Stanczuk, G. A., Currie, H., Baxter, G., Foster, A., Gibson, L., Graham, C., & Cuschieri, K. (2015). Cobas 4800 HPV detection in the cervical, vaginal and urine samples of women with high-grade CIN before and after treatment. *Journal of Clinical Pathology, 68*(7), 567-570. doi:10.1136/jclinpath-2014-202851
- 14. Burroni, E., Bonanni, P., Sani, C., Lastrucci, V., Carozzi, F., & The Hpv Screevacc Working Group Ann. (2014). Human papillomavirus prevalence in paired urine and cervical samples in women invited for cervical cancer screening. *Journal of Medical Virology*, *87*(3), 508-515. doi:10.1002/jmv.24085
- 15. Hagihara, M., Yamagishi, Y., Izumi, K., Miyazaki, N., Suzuki, T., Kato, H., . . . Mikamo, H. (2016). Comparison of initial stream urine samples and cervical samples for detection of human papillomavirus. Journal of Infection and Chemotherapy, 22(8), 559-562. doi:10.1016/j.jiac.2016.05.009





- 16. Khunamornpong, S., Settakorn, J., Sukpan, K., Lekawanvijit, S., Katruang, N., & Siriaunkgul, S. (2016). Comparison of Human Papillomavirus Detection in Urine and Cervical Samples Using High-Risk HPV DNA Testing in Northern Thailand. Obstetrics and Gynecology International, 2016, 1-8. doi:10.1155/2016/6801491
- 17. Cuzick, J., Cadman, L., Ahmad, A. S., Ho, L., Terry, G., Kleeman, M., . . . Erlander, M. G. (2017). Performance and Diagnostic Accuracy of a Urine-Based Human Papillomavirus Assay in a Referral Population. Cancer Epidemiology Biomarkers & Prevention, 26(7), 1053-1059. doi:10.1158/1055-9965.epi-16-0960
- 18. Nilyanimit, P., Chansaenroj, J., Karalak, A., Laowahutanont, P., Junyangdikul, P., & Poovorawan, Y. (2017). Comparison of human papillomavirus (HPV) detection in urine and cervical swab samples using the HPV GenoArray Diagnostic assay. *PeerJ*, 5. doi:10.7717/peerj.3910
- 19. Lee, H., Choi, M., Hwang, S., & Cho, Y. (2018). A Versatile Nanowire Platform for Highly Efficient Isolation and Direct PCR-free Colorimetric Detection of Human Papillomavirus DNA from Unprocessed Urine. Theranostics, 8(2), 399-409. doi:10.7150/thno.21696
- 20. Vergara, N., Balanda, M., Hidalgo, W., Martín, H. S., Aceituno, A., Roldán, F., . . . Ramírez, E. (2017). Detection and genotyping of HPV in urine samples from Chilean women attending primary health care centers. *Medical Microbiology and Immunology*. doi:10.1007/s00430-017-0530-
- 21. 24.Leeman, A., Pino, M. D., Molijn, A., Rodriguez, A., Torné, A., Koning, M. D., . . . Quint, W. (2017). HPV testing in first-void urine provides sensitivity for CIN2 detection comparable with a smear taken by a clinician or a brush-based self-sample: cross-sectional data from a triage population. *BJOG: An International Journal of Obstetrics & Gynaecology, 124*(9), 1356-1363. doi:10.1111/1471-0528.14682
- 22. Lorenzi, A. T., Fregnani, J. H., Dockter, J., Fitzgerald, K., Strohecker, E., Eaton, B., . . . Longatto-Filho, A. (2018). High-Risk Human Papillomavirus Detection in Urine Samples From a Referral Population With Cervical Biopsy-Proven High-Grade Lesions. Journal of Lower Genital Tract Disease, 22(1), 17-20. doi:10.1097/lgt.00000000000352
- 23. Sahasrabuddhe, V. V., Gravitt, P. E., Dunn, S. T., Robbins, D., Brown, D., Allen, R. A., . . . Wentzensen, N. (2014). Evaluation of clinical performance of a novel urine-based HPV detection assay among women attending a colposcopy clinic. *Journal of Clinical Virology, 60*(4), 414-417. doi:10.1016/j.jcv.2014.04.016
- 24. Sahasrabuddhe, V. V., Gravitt, P. E., Dunn, S. T., Brown, D., Allen, R. A., Eby, Y. J., . . . Wentzensen, N. (2013). Comparison of Human Papillomavirus Detections in Urine, Vulvar, and Cervical Samples from Women Attending a Colposcopy Clinic. Journal of Clinical Microbiology, 52(1), 187-192. doi:10.1128/jcm.01623-13
- 25. Mendez, K., Romaguera, J., Ortiz, A. P., López, M., Steinau, M., & Unger, E. R. (2013). Urine-based human papillomavirus DNA testing as a screening tool for cervical cancer in high-risk women. International Journal of Gynecology & Obstetrics, 124(2), 151-155. doi:10.1016/j.ijgo.2013.07.036
- 26. Piyathilake, C. J., Badiga, S., Chambers, M. M., Brill, I. K., Matthews, R., & Partridge, E. E. (2016). Accuracy of urinary human papillomavirus testing for the presence of cervical human papillomaviruses and higher grades of cervical intraepithelial neoplasia. *Cancer, 122*(18), 2836-2844. doi:10.1002/cncr.30123
- 27. Liu, T., Xie, R., Luo, L., Reilly, K. H., He, C., Lin, Y., ... Wang, H. (2014). Diagnostic validity of human papillomavirus E6/E7 mRNA test in cervical cytological samples. Journal of Virological Methods, 196, 120-125. doi:10.1016/j.jviromet.2013.10.032
- 28. Asciutto, K. C., Ernstson, A., Forslund, O., & Borgfeldt, C. (2018). Self-sampling with HPV mRNA analyses from vagina and urine compared with cervical samples. Journal of Clinical Virology, 101, 69-73. doi:10.1016/j.jcv.2018.02.002
- 29. Park, S., Wang, H., Kim, S., Kim, G., Bong, S., Jang, H., . . . Lee, D. (2016). Performance of HPV E6/E7 mRNA Genotyping Test on Paired Cervical Cancer Exfoliated Cells and Formalin Fixed Paraffin Embedded Tissues. *Biomedical Science Letters*, 22(3), 98-106. doi:10.15616/bsl.2016.22.3.98
- 30. Zappacosta, R., Sablone, F., Pansa, L., Buca, D., Buca, D., & Rosini, S. (2017). Analytic and Diagnostic Performances of Human Papillomavirus E6/E7 mRNA Test on up-to 11-Year-Old Liquid-Based Cervical Samples. A Biobank-Based Longitudinal Study. International Journal of Molecular Sciences, 18(7), 1480. doi:10.3390/ijms18071483



Improving Lives Through the Prevention & Treatment of Anogenital & HPV-Related Diseases